

Evaluation of alternative methods to stem injections to apply phosphonate to avocado trees for *Phytophthora* control, *i.e.* bark and soil penetrants to enhance phosphonate uptake – Preliminary report

JJ Serfontein and J Wright

QMS Agri Science
PO Box 416, Letsitele 0885, South Africa
www.agriscience.co.za

ABSTRACT

Phytophthora cinnamomi has a wide host range and causes necrosis of roots and necrotic lesions in the trunk and stem, leading to shoot dieback or crown death. Chemical control of root rot with phosphite is achieved by the direct fungistatic action and/or an indirect mechanism of action. Stem injections have been shown to be effective application of phosphite, yet literature indicates that foliar bark sprays and soil drenches with the addition of penetrants can also be effective against root rot. The aim of this study was to compare the efficacy of phosphonate/bark penetrant trunk application (spray application), as well as soil drench to that of trunk injections for the control of avocado root rot. The trial was conducted in an 11-year-old 'Hass' avocado orchard in Politsi. Soil samples collected from the trial block tested positive for *Phytophthora* spp. Trees were rated before the commencement of treatments and again 18 weeks later. Seven treatments, replicated eight times, were applied at six week intervals. Initial tree ratings ranged from 4.25 to 5.25 and after 18 weeks the untreated control was the only treatment where tree rating increased, whereas all other phosphonate treatments improved tree health, with rating values decreasing between 0.125 and 0.875. This trial will continue until the end of season one and will be repeated the following season (August/September 2013). Tree ratings will be done after the first season and prior to the second season.

OBJECTIVES

To compare the efficacy of phosphonate/bark penetrant trunk application (spray application) as well as soil drench to that of trunk injections for the control of avocado root rot under South African conditions.

INTRODUCTION

Phytophthora cinnamomi is a soil- and water-borne plant pathogen with a wide host range throughout the world. *Phytophthora cinnamomi* invades the roots and/or collars of its hosts, causing symptoms such as necrosis of roots, cankers and necrotic lesions in the trunk and stem, which often leads to shoot dieback or crown death (Zentmyer, 1980).

Management of this disease relies heavily on chemical control, namely with phosphite (H₃PO₃), a neutralised solution of the phosphonate anion (Fenn & Coffey, 1984). H₃PO₃ is not metabolised and remains in the plant tissue for a considerable time, months to years depending on the plant species (Guest & Grant, 1991). Invading *P. cinnamomi* myce-

lium may be inhibited by the direct fungistatic action of H₃PO₃, yet, H₃PO₃ concentrations found in plant tissues are often well below concentrations found to be fungistatic *in vitro*, thus an indirect mechanism of action must also be invoked (Guest & Grant, 1991).

A study done by Tynan *et al.* (2001) indicated that foliar applications of phosphite remained effective for five to 24 months in native Australian plant spp. and Shearer & Fairman (2007) showed that when *Banksia* spp. were treated with phosphite by stem injections or foliar sprays, phosphite effectiveness persisted two years for foliar applications and four years for stem injections. Similar observations with cherry trees by Wicks & Hall (1988) indicated that the foliar sprays were not as persistent as stem injections, yet



they concluded that foliar sprays were less phytotoxic and were the most economical means for phosphite applications. Fosetyl-Al and potassium phosphonate applied to avocado trees as foliar sprays, has also indicated prolonged effective levels up to eight weeks (Ouimette & Coffey, 1989).

The use of bark penetrants in combination with phosphonates increases the uptake of the chemical significantly in woody plant species (McComb *et al.*, 2008). Garbelotto *et al.* (2007) also showed that the bark applications on coastal oak, to control sudden oak death, were effective only when a bark penetrant was added. The effect of silicon on *Phytophthora* root rot varies and it is not recommended in Australia for this purpose (Australian Avocado Industry Report, 2005 – 2006), whereas Bekker *et al.* (2007) found potassium silicate to have a positive effect on *Phytophthora* root rot control during dryer periods.

The following was written in the Australian Industry Report regarding the application method: "After the discovery that injection of trees with phosphorous acid can inhibit feeder root growth if applied at the commencement of root flush, we compared injections with trunk sprays for control of root rot. When injected, most of the phosphorous acid travels down to the roots. The concentration in the roots is relatively high and, therefore, inhibitory. When sprayed onto the trunks, a lower but more consistent supply to the roots, with little or none was ending up in the canopy. Levels in the roots are sufficient to see recovery in severely affected trees." – Australian Avocado Industry Report, 2005 – 2006.

The efficacy of soil drench applications as found by QMS and the minimal labour involved also prompted an interest in registration of phosphonates as soil drench *via* irrigation. As mentioned, this practice is already in place in some production areas and has been done for many years with success by some citrus producers. A fear exist that *Phytophthora* will become resistant against phosphonates if applied as a soil drench (Lucas McClain, personal communication), either directly or indirectly by less induced resistance. This possibility has been investigated by Dobrowolski *et al.* (2008) in Australia. Their results indicated that prolonged use of phosphonates in orchards does select isolates of *P. cinnamomi* less sensitive to phosphite *in planta*, as indicated by more extensive colonisation of phosphite treated plant tissue by isolates from orchards, than from strains

where phosphonates had never been used. However, the isolates used came from orchards where either stem injections or foliar applications have been done. The decrease in sensitivity was minor and *P. cinnamomi* has a low evolutionary potential. Whether soil application will enhance this potential is not known and has been discussed as early as 1997 (Weinert *et al.*, 1997).

In our earlier work with phosphonates on avocado nursery trees in bags, we found the Ammonium Phosphonate superior to Potassium Phosphonate products as a soil drench and it is also suggested by the suppliers as a soil drench (Dr Steve Engelbrecht, personal communication). Soil application rates will be based on those used in citrus where the dosage and number of applications per annum is based on canopy size. The maximum application rate for a 200 g a.i. per L will not exceed 62 g/tree (old big trees), unlike the rate of 2640 g/m² as suggested by Kaiser and Whiley (1998).

Literature shows that phosphonates can be applied to avocado trees effectively as a surface trunk spray or soil drench to control *Phytophthora* root rot when mixed with an appropriate penetrant, thus without the negative effects of trunk injections. This may have huge financial benefits to the South African avocado industry.

MATERIALS AND METHODS

The trial was conducted in an avocado orchard in Politsi. The orchard consisted of 11-year-old 'Hass' trees that showed signs of decline. Soil samples were collected from the trial block and were tested for the presence of *Phytophthora* spp. by using the soil bait test. Trees were rated before the commencement of treatments and again prior to the second trunk injection (18 weeks later). Single tree plots were randomised throughout the trial sight. Each of the seven treatments (Table 1) was replicated eight times. The first round of applications commenced the 10th of October 2012. Bark sprays and soil drench were applied at six week intervals (rain dependant).

RESULTS AND DISCUSSION

Soil collected from the trial block tested positive for the presence of *Phytophthora* spp.. Trees were initially rated on the 8th of October 2012, before treatments began. Trees were rated 18 weeks later (Preliminary results) (Fig. 1). Initial tree ratings ranged

Table 1. Treatments and dosage rates applied throughout the trial block.

Treatment number	Treatment description	Product	Application method	Active ingredient (ml/tree)
1	Untreated control	-	-	-
2	Trunk injections	Avoguard	Injections	3 x 5 ml
3	Brilliant (1X) + Link (1X)	Brilliant 300SL + Link	Bark spray	17 + 0.3 in 300 ml water
4	Brilliant (2X) + Link (2X)	Brilliant 300SL + Link	Bark spray	34 + 0.6 in 300 ml water
5	Brilliant (1X) + AnnGro (1X)	Brilliant 300SL + AnnGro	Bark spray	17 + 0.7 in 300 ml water
6	Brilliant (1X) + AnnGro (1X)	Brilliant 300SL + AnnGro	Soil drench	24 + 1 in 10 L water
7	Brilliant (1X) + FoliarComplex	Brilliant 300SL + FoliarComplex	Soil drench	24 + 4.8 in 10 L water



from 4.25 to 5.25 (all statistics to be done on completion of the trial). After 18 weeks tree improvement/decline could be observed. The untreated control was the only treatment where tree rating increased (from 4.63 to 4.88). All other treatments showed that phosphonate applications (injections, bark sprays and soil drenches) improved tree health, with rating values decreasing between 0.125 and 0.875 (Fig. 2).

Treatments will continue until the end of season one (April 2013) and will commence for season two after harvest (August/September 2013). Tree ratings will be done after the first season and prior to the second season.

ACKNOWLEDGEMENT

We thank the South African Avocado Growers' Association for financially supporting this project.

REFERENCES

BEKKER, T.F., LABUSCHAGNE, N., AVELING, T. & KAISER, C. 2007. Efficacy of water soluble potassium silicate against *Phytophthora* root rot of avocado under field conditions. *South African Avocado Growers' Association Yearbook* 30: 39-38.
 DOBROWOLSKI, M.P., SHEARER, B.L., COLQUHOUN,

I.J., O'BRIEN, P.A. & HARDY, G.E.StJ. 2008. Selection for decreased sensitivity to phosphite in *Phytophthora cinnamomi* with prolonged use of fungicide. *Plant Pathology* 57: 928-936.

FENN, M.E. & COFFEY, M.D. 1984. Studies on the *in vitro* antifungal activity of Fosetyl-Al and phosphorous acid. *Disease Control and Pest Management* 74: 606-11.

GARBELOTTO, M., SCHMIDT, D.J. & HARNIK, T.Y. 2007. Phosphite Injections and Bark Application of Phosphite + Pentrabark™ Control Sudden Oak Death in Coast Live Oak. *Arboriculture & Urban Forestry* 33(5): 309-317.

GIBLIN, F., PEGG, K., THOMAS, G., WHILEY, A., ANDERSON, J. & SMITH L. 2007. Phosphonate trunk injections and bark sprays. Proceedings VI World Avocado Congress (Actas VI Congreso Mundial del Aguacate) 2007. Viña Del Mar, Chile. 12-16 Nov. 2007. ISBN No 978-956-17-0413-8.

GIBLIN, F., PEGG, K., WILLINGHAM, S., ANDERSON, J., COATES, L., COOKE, T., DEAN, J. & SMITH, L. 2005. *Phytophthora* revisited. New Zealand and Australia Avocado Growers' Conference 2005. 20-22 September 2005. Tauranga, New Zealand.

GUEST, D. & GRANT, B. 1991. The complex action

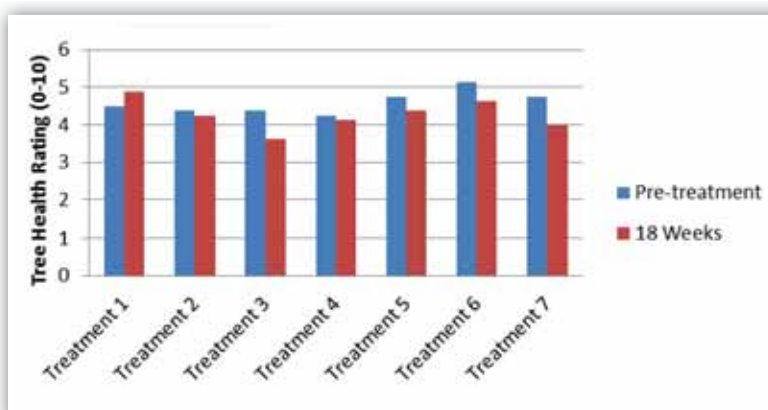


Figure 1. Comparison of tree health ratings before treatments and 18 weeks into the treatment program.

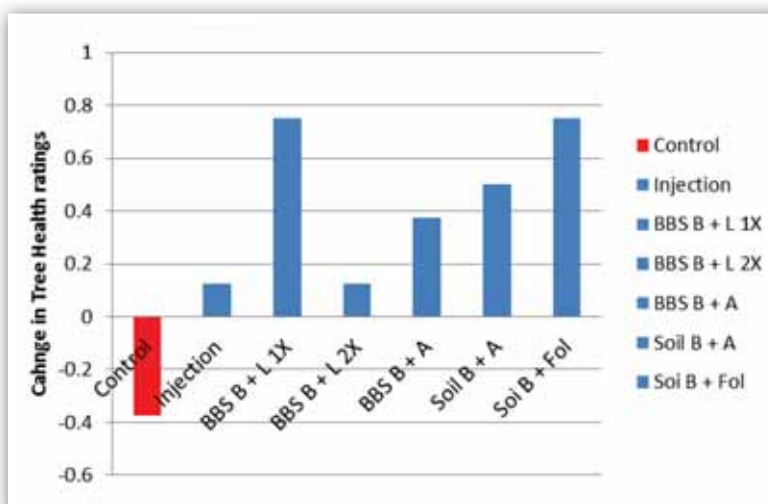


Figure 2. Change in tree health from beginning of season to 18 weeks into the phosphonite treatments.



- of phosphonates as antifungal agents. *Biological Reviews of the Cambridge Philosophical Society* 66: 195-87.
- KAISER, C. & WHILLEY, A.W. 1998. Effects of phosphonate soil drenching on avocados. *Talking Avocados*. 9(1): 15.
- MCCOMB, J.A., O'BRIEN, P., CALVER, M., STASKOWSKI, P., JARDINE, N., ESHRAGHI, L., ELLERY, J., GILOVITZ, J., SCOTT, P., O'BRIEN, J., O'GARA, E., HOWARD, K., DELL, B. & HARDY, G.E.StJ. 2008. Research into natural and induced resistance in Australian native vegetation of *Phytophthora cinnamomi* and innovative methods to contain and/or eradicate within localised incursions in areas of high biodiversity in Australia. Enhancing the efficacy of phosphite with the addition/supplementation of other chemicals such as those known to be involved in resistance. Prepared by the Centre for Phytophthora Science and Management for the Australian Government Department of the Environment, Water, Heritage and the Arts.
- OUIMETTE, D.G. & COFFEY, M.D. 1989. Phosphonate levels in avocado (*Persea americana*) seedlings and soil following treatment with fosetyl-Al or potassium phosphonate. *Plant Disease* 73: 212-5.
- SHEARER, B.L. & FAIRMAN, R.G. 2007. Application of phosphite in a high-volume foliar spray delays and reduces the rate of mortality of four *Banksia* species infected with *Phytophthora cinnamomi*. *Australian Plant Pathology* 36: 358-68.
- SMITH, L.A., DANN, E.K., PEGG, K.G., WHILEY, A.W., GIBLIN, F.R., DOOGAN, V. & KOPITKE, R. 2011. Field assessment of avocado rootstock selections for resistance to Phytophthora root rot. *Australasian Plant Pathology* 40: 39-47.
- TYNAN, K.M., WILKINSON, C.J., HOLMES, J.M., DELL, B., COLQUHOUN, I.J., MCCOMB, J.A. & HARDY, G.E.J. 2001. The long-term ability of phosphite to control *Phytophthora cinnamomi* in two native plant communities of Western Australia. *Australian Journal of Botany* 49: 761-70.
- WEINERT, M.P., DRENTH, A., SOO, S.H., IRWIN, J.A.G. & PEGG, K.G. 1997. Different phosphorous acid sensitivity levels in *Phytophthora cinnamomi* isolates from treated and untreated avocado trees. In: Proceedings of Australasian Plant Pathology Society, 11th Biennial Conference. p. 35.
- WICKS, T.J. & HALL, B. 1988. Preliminary evaluation of phosphorous acid, fosetyl-Al and metalaxyl for controlling *Phytophthora cambivora* on almond and cherry. *Crop Protection* 7: 314-8.
- ZENTMYER, G.A. 1980. *Phytophthora cinnamomi* and the diseases it causes. St. Paul, MN, USA: APS Press.

